Monday, February 24

Modeling Counts

Example: Consider the following data.

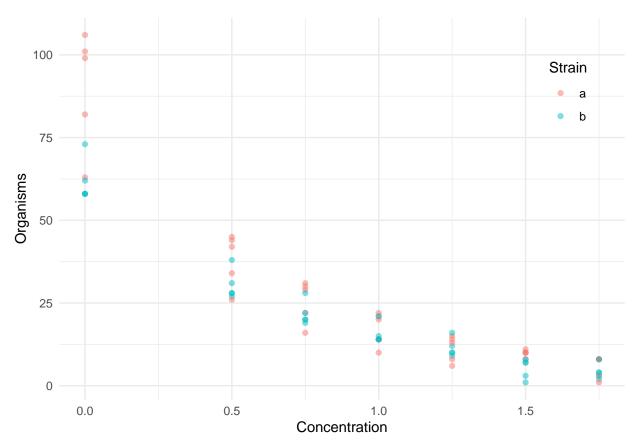
```
library(trtools) # for ceriodaphniastrain data
ceriodaphniastrain$strainf <- factor(ceriodaphniastrain$strain,
    levels = c(1,2), labels = c("a","b"))
head(ceriodaphniastrain)</pre>
```

	count	concentration	strain	strainf
1	82	0	1	a
2	58	0	2	b
3	106	0	1	a
4	58	0	2	b
5	63	0	1	a
6	62	0	2	b

tail(ceriodaphniastrain)

	count	concentration	${\tt strain}$	strainf
65	3	1.75	1	a
66	2	1.75	2	b
67	8	1.75	1	a
68	8	1.75	2	b
69	1	1.75	1	a
70	4	1.75	2	b

```
p <- ggplot(ceriodaphniastrain, aes(x = concentration, y = count, color = strainf)) +
geom_point(alpha = 0.5) + theme_minimal() +
theme(legend.position = "inside", legend.position.inside = c(0.9, 0.8)) +
labs(x = "Concentration", y = "Organisms", color = "Strain")
plot(p)</pre>
```



What are the complications when the response variable is a count?

- 1. Nonlinear models may be necessary because $E(Y_i) > 0$.
- 2. Heteroscedasticity because $Var(Y_i)$ tends to increase with $E(Y_i)$.
- 3. Non-normal discrete distribution.

One solution would be to use a *nonlinear* regression model combined with some method to account for the heteroscedasticity, and we will revisit this approach, but for now we will consider instead a specialized model that assumes a *Poisson* rather than a normal distribution of Y_i .

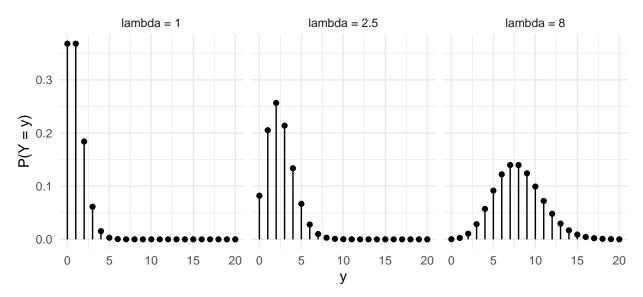
Poisson Regression

A random variable Y has a Poisson distribution if

$$P(Y=y) = \frac{\lambda^y e^{-\lambda}}{y!},$$

where y is a non-negative integer (i.e., y = 0, 1, 2, ...) and $\lambda > 0$ is the parameter of the distribution. Also note that y! is the *factorial* of y, defined as $y! = y \times (y - 1) \times (y - 2) \times \cdots \times 2 \times 1$ and 0! = 1.

It can be shown that if Y has a Poisson distribution then $E(Y) = \lambda$ and $Var(Y) = \lambda$. The parameter λ is sometimes called a "rate" parameter.



A regression model can be specified for a response variable with a Poisson distribution by assuming that

$$P(Y_i = y) = \frac{\lambda_i^y e^{-\lambda_i}}{y!}$$

where λ_i is a function of $x_{i1}, x_{i2}, \ldots, x_{ik}$. Since $\lambda_i > 0$ we might use

$$\lambda_i = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}).$$

This implies the *nonlinear* regression model

$$E(Y_i) = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}),$$

which can also be written as

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}$$

This kind of model is sometimes called a *log-linear* model. And because $Var(Y_i) = E(Y_i)$ the model assumes a certain pattern of heteroscedasticity. This kind of regression model is called a *Poisson regression* model.

Generalized Linear Models

Poisson regression is a member of a family of models known as *generalized linear models* (GLM). A generalized linear model has the form

$$g[E(Y_i)] = \underbrace{\beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik}}_{\eta_i}$$

where g is the link function and η_i is the linear predictor or systematic component. The link function is invertable so that we can also write

$$E(Y_i) = g^{-1}(\underbrace{\beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik}}_{\eta_i}) = g^{-1}(\eta_i).$$

Example: A linear regression model can be written as

$$E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik},$$

which implies that the link function is the "identity" function g(u) = u, and thus so is the inverse link function $g^{-1}(v) = v$.

Example: A Poisson regression model can be written as

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_1 x_{i2} + \dots + \beta_k x_{ik},$$

so the link function is $g(u) = \log(u)$, and the inverse link function is the exponential function $g^{-1}(v) = \exp(v)$, also written as e^{v} .

In a GLM the variance of Y_i is

$$\operatorname{Var}(Y_i) = \phi V[E(Y_i)]$$

where ϕ is a dispersion parameter and V is the variance function. That is, the variance of Y_i is proportional to some function of $E(Y_i)$.

Example: Linear models typically assume homoscedasticity meaning that $\operatorname{Var}(Y_i) = \sigma^2$ is a constant. Here the dispersion parameter is $\phi = \sigma^2$, and the variance function is just $V[E(Y_i)] = 1$.

Example: In Poisson regression we have that $Var(Y_i) = E(Y_i)$. Here the dispersion parameter is $\phi = 1$, and the variance function is the identity function $V[E(Y_i)] = E(Y_i)$.

We can write a GLM concisely as

$$E(Y_i) = g^{-1}(\eta_i),\tag{1}$$

$$\operatorname{Var}(Y_i) = \phi V[g^{-1}(\eta_i)], \tag{2}$$

where again

Temp

InsulAfter:Temp

$$\eta_i = \beta_0 + \beta_1 x_{i1} + \dots + \beta_k x_{ik},$$

to define the *mean structure* and a *variance structure* for Y_i , respectively. In general, specification of a GLM involves specifying three component parts:

- 1. The linear predictor $\eta_i = \beta_0 + \beta_1 x_{i1} + \dots + \beta_k x_{ik}$.
- 2. The link function g to relate $E(Y_i)$ to η_i as $g[E(Y_i)] = \eta_i$.
- 3. The distribution of Y_i , or the variance structure $\phi V[g^{-1}(\eta_i)]$.

The choice of distribution implies a certain variance structure when using glm. The distribution comes from a family of distributions known as the *exponential family* (not to be confused with what is called an *exponential distribution*).

Example: Normal linear regression is a GLM where g(u) = u, $g^{-1}(v) = v$, $\phi = \sigma^2$, and V(u) = 1 so that

$$E(Y_i) = \eta_i,\tag{3}$$

$$\operatorname{Var}(Y_i) = \sigma^2. \tag{4}$$

Example: Poisson regression is a GLM where $g(u) = \log(u)$, $g^{-1}(v) = e^v$, $\phi = 1$, and V(u) = u so that

$$\log E(Y_i) = \eta_i,\tag{5}$$

$$\operatorname{Var}(Y_i) = \exp(\eta_i). \tag{6}$$

In R the function glm can be used to specify a generalized linear model.

Example: Recall the model for the whiteside data.

-0.3932

0.1153

```
library(MASS)
m <- lm(Gas ~ Insul + Temp + Insul:Temp, data = whiteside)
summary(m)$coefficients

Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.8538 0.13596 50.409 7.997e-46
InsulAfter -2.1300 0.18009 -11.827 2.316e-16
```

0.03211

3.591 7.307e-04

0.02249 -17.487 1.976e-23

```
m <- glm(Gas ~ Insul + Temp + Insul:Temp, data = whiteside,
    family = gaussian(link = identity))
summary(m)$coefficients
```

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	6.8538	0.13596	50.409	7.997e-46
InsulAfter	-2.1300	0.18009	-11.827	2.316e-16
Temp	-0.3932	0.02249	-17.487	1.976e-23
InsulAfter:Temp	0.1153	0.03211	3.591	7.307e-04

Note that we do not explicitly state the variance structure (although we could — more on that later). Here the variance structure is implied by the choice of distribution.

Example: Now consider the following Poisson regression model for the ceriodaphniastrain data.

```
m <- glm(count ~ concentration + strainf, data = ceriodaphniastrain,
  family = poisson(link = log))
summary(m)$coefficients
```

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	4.455	0.03914	113.819	0.000e+00
concentration	-1.543	0.04660	-33.111	2.057e-240
strainfb	-0.275	0.04837	-5.684	1.313e-08

This model can be written as

 $E(Y_i) = \exp(\beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}),$

or

$$\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}$$

where x_{i1} is the concentration for the *i*-th observation, and x_{i2} is an indicator variable for strain such that

$$x_{i2} = \begin{cases} 1, & \text{if the strain is b} \\ 0, & \text{otherwise,} \end{cases}$$

so that the model can be written case-wise as

$$\log E(Y_i) = \begin{cases} \beta_0 + \beta_1 c_i, & \text{if the strain is a,} \\ \beta_0 + \beta_2 + \beta_1 c_i, & \text{if the strain is b,} \end{cases}$$

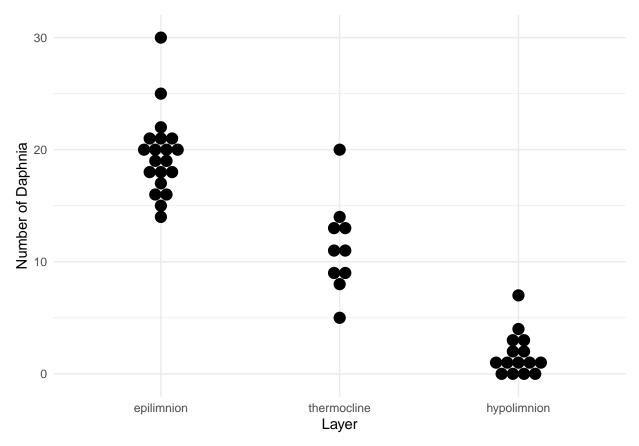
or

$$E(Y_i) = \begin{cases} \exp(\beta_0 + \beta_1 c_i), & \text{if the strain is a,} \\ \exp(\beta_0 + \beta_2 + \beta_1 c_i), & \text{if the strain is b,} \end{cases}$$

if we let $c_i = x_{i1}$ denote the concentration for the *i*-th observation. Also note that because Y_i is assumed to have a Poisson distribution, $Var(Y_i) = E(Y_i)$.

Example: Recall the daphnia survey.

```
library(trtools) # for daphniastrat
library(ggplot2)
p <- ggplot(daphniastrat, aes(x = layer, y = count)) +
  geom_dotplot(binaxis = "y", binwidth = 1, stackdir = "center") +
  labs(x = "Layer", y = "Number of Daphnia") + theme_minimal()
plot(p)
```



A Poisson regression model for these data might be specified as follows.

m <- glm(count ~ layer, family = poisson(link = log), data = daphniastrat)
summary(m)\$coefficients</pre>

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	2.9704	0.05064	58.661	0.000e+00
layerthermocline	-0.5456	0.10683	-5.107	3.272e-07
layerhypolimnion	-2.4204	0.20255	-11.950	6.519e-33

So the model is $\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2}$, where Y_i is the *i*-th count, and x_{i1} and x_{i2} are defined as

$$x_{i1} = \begin{cases} 1, & \text{if the } i\text{-th observation is from the thermocline layer,} \\ 0, & \text{otherwise,} \end{cases}$$

and

$$x_{i2} = \begin{cases} 1, & \text{if the } i\text{-th observation is from the hypolimnion layer,} \\ 0, & \text{otherwise.} \end{cases}$$

So the model can be written case-wise as

 $\log E(Y_i) = \begin{cases} \beta_0, & \text{if the } i\text{-th observation is from the epilimnion layer,} \\ \beta_0 + \beta_1, & \text{if the } i\text{-th observation is from the thermocline layer,} \\ \beta_0 + \beta_2, & \text{if the } i\text{-th observation is from the hypolimnion layer,} \end{cases}$

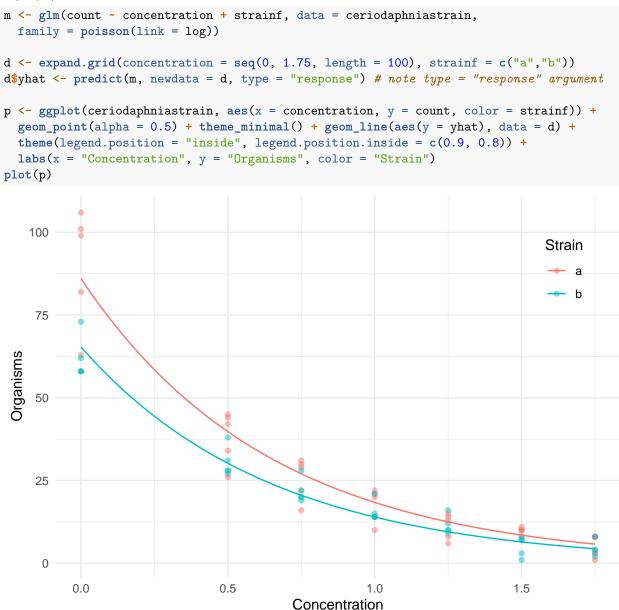
or

$$E(Y_i) = \begin{cases} \exp(\beta_0), & \text{if the } i\text{-th observation is from the epilimnion layer,} \\ \exp(\beta_0 + \beta_1), & \text{if the } i\text{-th observation is from the thermocline layer,} \\ \exp(\beta_0 + \beta_2), & \text{if the } i\text{-th observation is from the hypolimnion layer.} \end{cases}$$

And of course we have that $Var(Y_i) = E(Y_i)$.

Visualization of a GLM

Visualization of a GLM be done in the usual way provided we use the type = response option when using predict. The default, which is type = link, returns $\hat{\eta}_i = \hat{\beta}_0 + \hat{\beta}_1 x_{i1} + \cdots + \hat{\beta}_k x_{ik}$ which is the estimate of $\log E(Y_i)$.



To compute confidence intervals for the expected response you can use the glmint function from the trtools package.

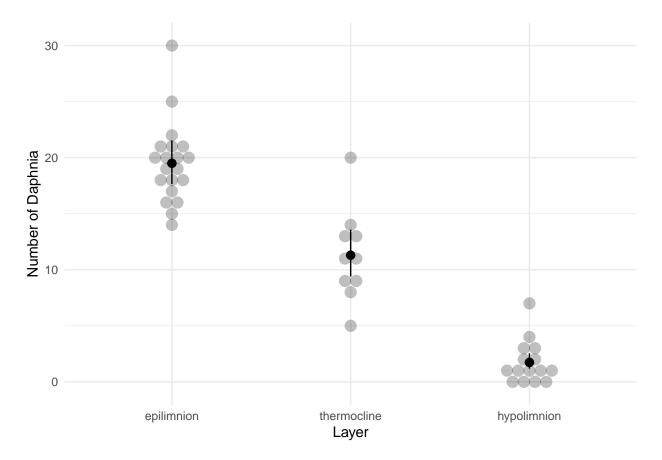
```
d <- expand.grid(concentration = seq(0, 1.75, length = 100), strainf = c("a","b"))
d <- cbind(d, glmint(m, newdata = d))
head(d)</pre>
```

	concentration	strainf	fit	low	upp
1	0.00000	a	86.03	79.67	92.88
2	0.01768	a	83.71	77.60	90.30
3	0.03535	a	81.46	75.58	87.79

```
a 79.27 73.61 85.35
4
        0.05303
                       a 77.13 71.69 82.99
5
        0.07071
        0.08838
                       a 75.06 69.82 80.68
6
p <- ggplot(ceriodaphniastrain, aes(x = concentration, y = count, color = strainf)) +
  geom_point(alpha = 0.5) + theme_minimal() +
  geom_line(aes(y = fit), data = d) +
  geom_ribbon(aes(ymin = low, ymax = upp, y = NULL, fill = strainf),
    color = NA, alpha = 0.25, data = d) +
  theme(legend.position = "inside", legend.position.inside = c(0.9, 0.8)) +
  guides(fill = "none") +
  labs(x = "Concentration", y = "Organisms", color = "Strain")
plot(p)
   100
                                                                                  Strain
                                                                                   🗕 а
                                                                                      b
     75
Organisms
     50
     25
     0
          0.0
                                0.5
                                                       1.0
                                                                             1.5
                                           Concentration
We might visualize the model for the daphniastrat data as follows.
```

```
m <- glm(count ~ layer, family = poisson(link = log), data = daphniastrat)</pre>
```

```
d <- data.frame(layer = unique(daphniastrat$layer))
d <- cbind(d, glmint(m, newdata = d))
p <- ggplot(daphniastrat, aes(x = layer, y = count)) +
   geom_dotplot(binaxis = "y", binwidth = 1, stackdir = "center", alpha = 0.25) +
   labs(x = "Layer", y = "Number of Daphnia") +
   theme_minimal() + geom_pointrange(aes(y = fit, ymin = low, ymax = upp), data = d)
plot(p)</pre>
```



GLMs Versus Nonlinear Regression

There is a very close relationship between a GLM and a nonlinear regression model. We can try to estimate the model above using nlm as follows, using the estimates as starting values for convenience.

Estimate Std. Error t value Pr(>|t|) b0 4.4821 0.02894 154.86 2.486e-87 b1 -1.5679 0.06232 -25.16 7.441e-36 b2 -0.3267 0.04506 -7.25 5.402e-10

The estimates are not the same. But consider that the GLM assumes that

 $\operatorname{Var}(Y_i) = E(Y_i),$

so our weights should be $w_i = 1/E(Y_i)$. Consider then an iteratively weighted least squares algorithm with $w_i = 1/\hat{y}_i$.

```
ceriodaphniastrain$w <- 1
for (i in 1:10) {
    m <- nls(count ~ exp(b0 + b1 * concentration + b2 * (strainf == "b")),
    data = ceriodaphniastrain, start = list(b0 = 7, b1 = -2, b2 = 0.12),
    weights = w)
    ceriodaphniastrain$w <- 1 / predict(m)
}
summary(m)$coefficients</pre>
```

```
Estimate Std. Error t value Pr(>|t|)
b0 4.455 0.04272 104.273 7.167e-76
b1 -1.543 0.05087 -30.334 7.309e-41
b2 -0.275 0.05280 -5.208 1.988e-06
```

Compare that with what we obtained using glm.

m <- glm(count ~ concentration + strainf, data = ceriodaphniastrain, family = poisson(link = log)) summary(m)\$coefficients

Estimate Std. Error z value Pr(>|z|)(Intercept)4.4550.03914113.8190.000e+00concentration-1.5430.04660-33.1112.057e-240strainfb-0.2750.04837-5.6841.313e-08

The parameter estimates are the same, but the standard errors are not. Why? The GLM assumes $Var(Y_i) = E(Y_i)$ whereas the iteratively weighted least squares approach assumes $Var(Y_i) \propto E(Y_i)$.

GLMs Versus Response Variable Transformations

It is important to note that, for example, the GLM

 $\log E(Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik}.$

is not equivalent to a linear model with a transformed response,

$$E(\log Y_i) = \beta_0 + \beta_1 x_{i1} + \beta_2 x_{i2} + \dots + \beta_k x_{ik},$$

because $\log E(Y_i) \neq E[\log Y_i]$ (although in practice they can produce similar results).